Phytochemical Investigation of EtOH Extract of Flowers of

Senecio Chrysanthemoide, Asteraceae *1S. C. Sati, ²Maneesha D. Sati, ³Shikha Saxena and ⁴I. P. Pandey *1Department of Chemistry, H.N.B. Garhwal University (A Central University) Srinagar Garhwal, Uttarakhand, India.246174 ²Department of Chemistry, Govt. Degree College Devprayag, TehriGarhwal, Uttarakhand, India ³Department of Chemistry, D.A.V. (PG) College Dehradun, Uttarakhand, India ⁴Professor Emeritus, Dehradun, Uttarakhand, India ***Email: sati_2009@rediff mail.com** DOI 10.51129/ujpah-2022-32-1(4)

Received –May 17, 2022 Revised – May 27, 2022 Accepted –June 1, 2022 Published – June 18, 2022

Abstract - From the ethanolic extract of flowers of *Seneciochrysanthemoide*one flavone diglycosideonexanthone glycoside have been isolated. The structures of the isolated compounds were identified with the help of chemical and spectral studies.

Key words: *Senecio Chrysanthemoide*, Asteraceae, Flavone Glycosides and Xanthone.

Introduction

Seneciospecies are used in folk medicine for the treatment of cuts, wounds and as antiemetic, anti-inflammatory, antimicrobialand vasodilator. The parts mostly used are leaves, stem and flowers¹, the pyrrolizidine alkaloids and the furanosesquiterpenoids are the most important constituents of this genus and thought to be responsible for all of pharmacological activities²⁻⁵. The genus Senecio, belongs to the tribe Senecioneae, is the largest and most complex genus in the family of the Asteraceae (Compositae) includes more than 1500 species with a worldwide distribution⁶. The chemical constituents of the genus *Senecio* include notably sesquiterpenoids, monoterpenoids⁷⁻⁸, diterpenoids⁹,triterpenoids¹⁰, phenolicand

flavonoidcompounds¹¹⁻¹⁶, essential oils¹⁷and pyrrolizidine alkaloids⁵. In India the genus Senecio comprises 43 species including S. chrysanthemoide which grows endemically in all over India¹⁸. The present paper deals with isolation and characterization of flavone di-glucoside and 3acetoxyxanthone from the alcoholic extract of the flowers of S. chrysanthemoide.

Material and Methods

Mps. Uncorrected, Columnchromatography was carried out on silica gel (60-120) mesh. Merck, eluting solvent (CHCl₃MeOH). U.V. was taken in MeOH. ¹H-NMR spectra were taken using TMS as internal standard and CDCl₃ and CD₃OD as solvents, all the signals are expressed as values downfield from TMS.

Collection of Plant Material

The Flowers of *S. chrysanthemoides*were collected in flowering stage from in Tunganath Chopta, Rudraprayag U.K. India. The plant was identified by expert Taxonomists Department of Botany HNB Garhwal University Srinagar Garhwal. The

voucher specimen (H.N.24446) is available in the herbarium of Plant Identification Laboratory department of Botany HNB Garhwal University Srinagar Garhwal.

Extraction and Isolation

The air-dried and coarsely powered flowers of the plant were defatted with light petroleum in a soxhlet. The defatted mass exhaustively extracted was repeatedly with 90% aqueous EtOH, until the extractive became colorless. All the extracts were mixed and concentrated under reduced pressure using rotatory vacuum evaporator.The concentrated extract was adsorbed on silica gel and through fractionated column chromatography using the solvent system chloroform: methanol (97:3). The polarity of solvent was gradually increased by addition of methanol. Repeated column chromatography afforded compounds 1 and 2 together with β -sitosterol and kaemferol.

Results and Discussion

The ethanolic extract of flowers of S. *chrysanthemoides* on repeated column chromatography over silica gel afforded compounds 1 and compound 2 together with β -sitosteroland kaempferol. The structure of β –sitosterol and kaempferol was confirmed by their comparison with an authentic sample (TLC) and reported data of the compound¹⁹. The structure of compound 1 was identified as α -L arabinopyranosyl (1 \rightarrow 3) β -D glucopyranosyl $(1 \rightarrow 3)\beta$ -hydroxyoleane-12-ene 28 methyl acetate and compound 2 as xanthone3-Acetoxy-1-hydroxy-6-

 $methoxy8\text{-}O\text{-}\beta\text{-}Dglucopyranosyl(1{\rightarrow}3)\alpha\text{-}$

L rhamnopyr-anosylwith the help of chemical and spectral studies.

Compound 1

It was obtained as crystalline solid from MeOH

 Melting Point
 : 220-222 ° C

 Molecular Formula
 : C₃₀ H₂₈O₁₆

 Molecular Weight
 : 644

¹H-NMR (CDCl₃,100MHz,δppm):2.66

(d, J=1.6 Hz), 3.49 (s), 3.25 (s), 7.70 (d, J=8.4 Hz), 6.38 (d, J=7.5 Hz),7.02 (d, J=7.5 Hz), 7.29 (d, J= 1.5 Hz), 5.45 (d, J=6.8 Hz), 5.49 (d, J=7.5 Hz),5.42 (d, J=6.8 Hz,C-1' anomeric proton),4.98 (d, J=5.8 Hz,C-1''), 3.65-4.72 (sugar multiplets)

¹³C-NMR(CDCl₃,150MHz, δ ppm):79.3 (C-2), 43.0 (C-3), 190.1(C-4) 162.8 (C-5), 110 (C-6), 163.5 (C-7), 99.7 (C-8),127.9 (C 9),115.7(C-10), 123.9 (C-11), 113.6 (C-12),141.1 (C-13), 147.0 (C-14), 41(-OCH₃), 40.2 (-OCH₃), 20.9 (-OCH₃)

Arabinopyranosyl: 101.9 (C-1'), 73.1 (C-2'), 76.0 (C-3'), 69.9 (C-4'), 76.0(C-5'), 69.4 (C-6')

Glucopyranosyl: 103.1 (C-1"), 73.1 (C-2"), 75.5 (C-3"), 69.3 (C-4") 75.0 (C-5"), 60.3 (C-6")

The compound 1 was found to be positive for coloration with methanolic FeCl₃ and Shinoda test Molish test (Mg/HCl)there by indicating flavonoid nature of compound. IR spectra of compound displayed absorption bands 3302, 1665, 1622, 1505 showed at presence of hydroxyl and carbonyl functions in the compound. The molecular formula of compound was calculated as which corres- pond the $C_{30}H_{28}O_{16}$ molecular weight 644 amu, due to the presence of molecular ion peak at m/z644[m]⁺,645[M+H]⁺, 675[M+CH₃OH]. Other fragment ion appeared peaks at m/z

peaks appeared at m/z452[(M+H)⁺-(OCH₃+162)] and 273[(M+H)⁺-(OCH₃+O

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 $H+2\times162$]⁺.The¹HNMR spectrum of compound displayed two doublets at δ 6.38 (d, J = 7.5 Hz , C-12) and δ 7.02 (d,J = 7.5Hz, C-13) and one doublet of 1.6 Hz coupling constant appeared at δ 2.66 (C-3) assigned for flavonoid proton. A doublet of 8.4 Hz at δ 7.70 ascribed to C-6 of flavonoid. The position of two, 3H proton singlet at δ 3.49 and δ 3.25 assigned for two OCH3 The ¹H signals appeared at δ 3.60-5.4 ascribes the sugar moiety in the molecule. The position of two doublets at δ 5.42 (d, J= 6.8 Hz) and δ 4.98 (J=7.5 Hz) represents two anomeric sugars. The presence of two sugars were further in agreement with the FAB-MS data displayed peaks at m/z 452 and 273

arose by the loss of two sugar molecule from the molecular ion peak.All These values were in agreement with its¹³C NMR data. The coupling constant value of anomeric sugar showed β configuration in both the sugar molecule. Compound when hydrolyzed with 7% methanolicHCl furnished two glucose molecules (from PC and TLC). Methylation, methanolysis and partial hydrolytic studies revealed the position of both the sugar in different carbon. All these data were in agreement with the reported data of flavonoidal glycoside²⁰.Hence compound was identified as Flavon 5, 4'dimethox 8 **7-O-β-D** methvl glucopyranoside-5'βDglucopyranoside. (Figure-1)

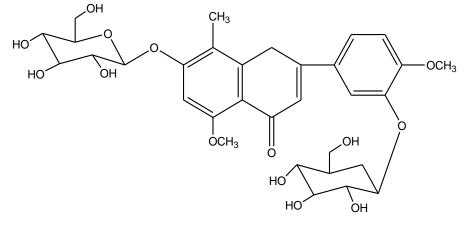


Figure – 1

Compound 2

It was obtainted as colourless crystalline solidfromMeOH

M.P.: 196-198⁰ C FAB MS (m/z): $606[M]^+$, $607 [M+H]^+$, $460 [M+ H-Rham]^+298 [MH-(rha+glu)]^+$, 255, 209¹HNMR (CDCl₃100MHz, δ ppm):6.83(d,J=8.4Hz), 7. 68(s), 7.24(s), 6.62(d, J=8.4Hz), 2.48 (s), 2.65(s), 9.06 (s) Glycone: 5.43 (d,J=7.8Hz), 5.2(s), 3.5-4.9 (Sugarproton). Molecular weight: 606I.R. (v_{max}^{KBr}) cm⁻¹: 2995, 3000, 1640

Molecular formula :C₂₈ H₃₀ O₁₅

¹³CNMR (CDCl₃150MHz, **δppm**):Aglycone163.9 (C-1), 117.6 (C-2),1473 (C-3),122.21(C-4),125.4(C-5),145(C-6),128.6(C-7),163.3(C-8),191.4(C-9), 18.4(C-10),138.7(C-11), 116.3 (C-12), 115 .7(C-13), 115.4(C-14), 176.0 (C=O). 43.3(CH3),56.3 (OMe) Glycone:103.0 (C-1'),73.6 (C-2'),79.6 (C-3'), 63.1 (C-4'), 76.3 (C-5'), 61.2(C-6').

Glycone:101.9 (C-1"),73.4 (C-2"),77.4(C-

It gave green coloration with FeCl₃ and also responded positive test with Molishreagent there by indicating phenolic nature of the compound²¹. IR spectrum of compound showed characteristic bands 2995, 3000 and 1640 cm⁻¹ for at phenolichydroxy and carbonyl groups. ¹H - NMR spectrum of the compound displayed two 1H proton singlets at δ 7.24 and δ 7.68 for C-5 and C-4 hydrogen, which confirmed the presence of xanthone skeleton in compound. Two doublets of 8.4 Hz coupling constant appeared at δ 6.83 and δ 6.62 showing an ortho-coupling. Two up field sharp δ 2.48 and δ 2.65 indicated singlets at the presence of two methoxy group in compound, whereas a weak singlet at δ 9.06 were assigned for hydroxyl group. Two anomeric proton resonated at δ 5.43 (d, J = 7.8 Hz) and δ 5.2 (s) with other sugar peaks appeared between δ 3.1-4.9 assigned for 10 sugar protons. The molecular weight of compound was deduced as 606 amu which corresponding the molecular formula C₂₈H₃₀O₁₅, due to the presence of molecular ion peak at m/z $607 [M+H]^{+}$ The presence of these different groups were in agreement with the mass fragmentation of a compound as shown by its FAB-MS which furnished peaks at $[M+H-Rham]^+$, m/z 460 298[M+H-

3"),69.9(C-4"),60.9(C-5"),18.5(C-6').

 $(Rham+Glu)]^+$, 255 $[M-(2Glu+COCH_3)]$ and 209 $[M-(2gly+COCH_3+OCH_3)]$. The structure of glycone

was further supported by its hydrolysis studies. Compound was hydrolyzed with 7% methanolicHCl for about 8 hours. It furnished an aglycone identified as 3-Acetoxy 1-hydroxy 6-methoxyxanth- one from its reported data. The neutralized hydrolysate gave two sugars identified as glucose (PC, Rf value 0.18) and rhamnose (PC, Rf value 0.37). Compound 2 on partial hydrolysis yield one rhamnose (PC, Rfvalue 0.37) and one Mono glycosidicaglycone. The prosapogenin on further HOH vield one xanthone as an aglycone (TLC) and glucose (PC, RF) showed the sequential loss of sugars.

The point of glycosidation was established by the¹³C-NMR data of sugars which was fixed at C-3 of glucose with C-1 of rhamnose. The configuration was found to be β in glucose and α in rhamnose by the J value of anomeric sugar in its ¹H-NMR spectrum. These all values were compared with the reported data of xanthone glycosides²⁰. Thus on the basis of spectral studies compound 2 was identified as **xanthone 3-Acetoxy-1-hydroxy-6-methoxy 8-O-\beta-D glucopyranosyl (1\rightarrow3) \alpha-L rhamnopyranoside. (Figure-2)**

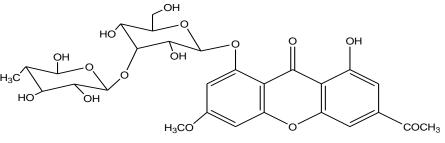


Figure – 2

Acknowledgement

Authors are thankful to IIT New Delhi for NMR spectra and CDRI Lucknow for Mass spectrum of compounds.

Disclaimer Statement

Authors declare that no competing interest exists. The products used for this researchare commonly used products in research. There is no conflict of interest between authors and producers of the product.

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